

## Effects of human growth hormone (hrGH) treatment on amine metabolism in rats subjected to extensive small bowel resection

W. A. Fogel<sup>1</sup>, K. Sasiak<sup>1</sup>, J. Socha<sup>2</sup>, and W. Andrzejewski<sup>1</sup>

<sup>1</sup>Institute of Biogenic Amines, Polish Academy of Sciences, Lodz, and

<sup>2</sup>Department of Gastroenterology & Nutrition, The Children's Memorial Health Institute, Warsaw, Poland

Received February 13, 2002

Accepted June 27, 2002

Published online November 14, 2002; © Springer-Verlag 2002

**Summary.** The effect of human recombinant growth hormone (hrGH) on intestinal adaptation in rats subjected to massive small bowel resection has been followed by monitoring changes in the tissue polyamine system and in red blood cell (RBC) polyamine levels. In parallel, the activities of monoamine oxidase A and B and diamine oxidase, the enzymes that catalyse one of the major routes of biogenic amine metabolism, oxidative deamination, were also examined.

The results suggest that whilst hrGH treatment accelerates adaptive intestinal hyperplasia evoked by the resection, it has no significant effect on RBC polyamine level or gut mucosal DNA concentration as measured 3 weeks post surgery. hrGH treated operated rats exhibited significantly lower amine oxidase activities which implies that GH may alter biogenic amine systems.

**Key words:** Growth hormone – Intestinal adaptation – Amine metabolism – Erythrocyte polyamines

### Introduction

Extensive small intestine resection results in the loss of absorptive surfaces, acceleration of intestinal transit and, as a consequence, in malnutrition, weight loss, diarrhoea and other complications of short bowel syndrome (Vanderhoof et al., 1992). The availability of human recombinant growth hormone (hrGH) and stimulatory effects of GH on gut growth (Leblond and Carriere, 1955; Lehy et al., 1986; Lobie et al., 1990) suggested its use in the treatment of short bowel syndrome (Ellegard et al., 1997). The trophic response of GI tract epithelium to hormones such as GH is mediated by polyamines, which are vital in cell proliferation (Janne et al., 1978). Tissue polyamine concentrations directly reflect growth stimulation or

retardation (Seidel, 1986; Hosomi et al., 1987). Likewise, it has been shown that measurement of an erythrocytic polyamine concentration may be diagnostically useful. Polyamines produced and secreted in excess by rapidly growing tissues enter the circulation where they are transported almost exclusively by red blood cells (RBC). Since these cells are unable to synthesise or degrade polyamines, the RBC polyamine level may serve as a marker of cellular proliferation (Moulinoux et al., 1981, 1984, 1991).

Recent studies have shown that in addition to its growth promoting effects, growth hormone exerts multiple metabolic effects. However, with the exception of some data on the polyamine system, there is little available information about amine metabolism.

This study was undertaken in rats to: 1/ evaluate the effects of hrGH by monitoring polyamine and amine metabolism parameters in the adapting short bowel and 2/determine whether erythrocyte polyamine concentrations reliably reflect the proliferative activity of the remaining bowel and may be suitable clinically to follow adaptation processes in children with short bowel syndrome.

### Materials and methods

All procedures strictly followed the Polish legislation concerning animal experiments and were approved by the local animal ethics committee.

A 70% resection of the small intestine of Wistar rats was performed under ether anaesthesia leaving equidistant lengths of bowel from pylorus and ileocecal valve. Two controls were employed: rats with only transected small intestine and intact rats.

Recombinant human GH (0.2IU, s.c., Saizen, Serono, Switzerland) was administered once daily for 5 or 10 days, to randomly selected rats from the second postoperative day onwards.

Animals were sacrificed by guillotine decapitation 8, 13 and 21 days after surgery. Trunk blood was collected for RBC polyamine measurements. Remnant small bowel was freed of attached mesentery, removed, its length and weight recorded for mucosal hyperplasia assessment. Distal ileum of a similar length was dissected from control rats and was handled in a similar manner. After thoroughly flushing with cold saline, the intestinal mucosa was scraped off. The liver, which is the highest source of monoamine oxidase enzymes, was also sampled. All tissue samples were frozen in liquid nitrogen and stored at  $-70^{\circ}\text{C}$  until assayed for enzyme activities and amine and polyamine concentrations.

#### Biochemical analyses

Ornithine decarboxylase was measured with L [1- $^{14}\text{C}$ ]-ornithine, using the  $^{14}\text{CO}_2$  trapping method (Kobayashi, 1963). Monoamine oxidase (MAO-A and MAO-B) activities were estimated in tissue homogenates with radioassays (Fowler and Tipton, 1982), employing serotonin (final concentration  $200\mu\text{M}$ ) or beta-phenylethylamine (final concentration  $20\mu\text{M}$ ) and specific inhibitors deprenyl and clorgyline ( $0.3\mu\text{M}$  each), respectively. Diamine oxidase activity was assayed with  $^{14}\text{C}$  putrescine (Fogel et al., 1985) and the polyamine oxidase against acetylspermine with homovanilic acid as a fluorogen (Matsumoto et al., 1984). All enzyme activities are expressed in pmol/min/mg protein.

*Polyamines, spermidine (SPD) and spermine (SPM)*, after extraction from erythrocytes or intestinal mucosa with 0.4M perchloric acid, were dansylated, separated by reversed phase HPLC using a LiChrosorb RP 18 ( $5\mu\text{m}$ ) column and quantitated by fluorometry as described by Mates et al., (1992). The concentration of polyamines in erythrocytes is given as nmol/ml of packed RBC, in tissue in nmol/g wet weight. Tissue histamine concentrations were determined by a fluorometric procedure after isolation on a small Cellex P column as described elsewhere (Fogel, 1988). DNA concentrations was assayed using the diphenylamine method (Burton, 1968). Protein was measured according to Lowry et al. (1951).

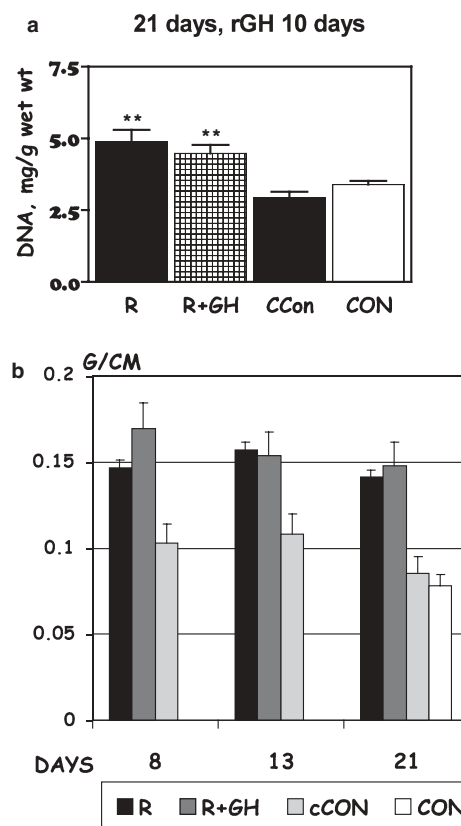
#### Statistical analysis

The values are presented as mean  $\pm$  SD. Differences between groups were assessed with paired or unpaired t-tests as appropriate.

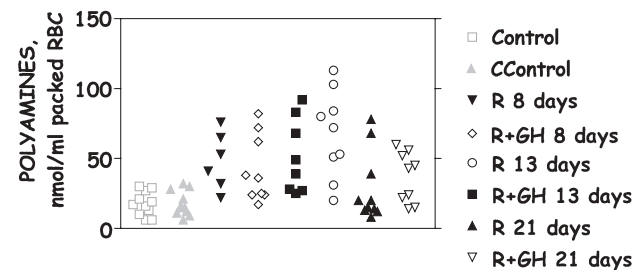
### Results

Resection was a powerful stimulus for regenerative processes in the intestine. DNA concentrations and the mucosal hyperplasia index: weight/length ratio were significantly higher in operated vs control rats ( $p < 0.01$  and  $p < 0.05$ , respectively). hrGH treatment, however, had no significant effect, although on 8<sup>th</sup> postoperative day, hormone-treated rat intestinal mucosa was somewhat thicker (Fig. 1a, b).

As shown in Fig. 2, red blood cell polyamine concentrations were significantly higher in operated versus control rats ( $p < 0.01$  at least), with the

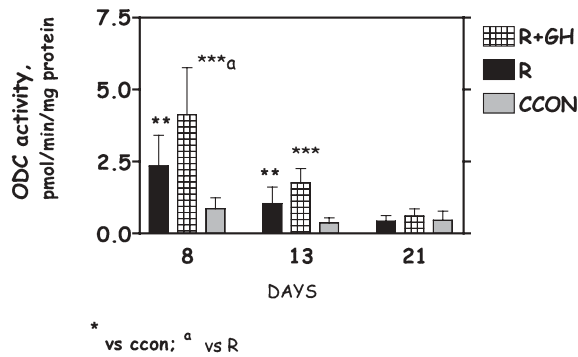


**Fig. 1.** Adaptive processes after massive small bowel resection; the effect of growth hormone treatment. Panel **a**: Concentration of mucosal DNA. Panel **b**: mucosal hyperplasia index: weight/length ratio. R, resection; R + GH, resection and growth hormone treatment; Ccon, bowel transection control; CON, intact rats

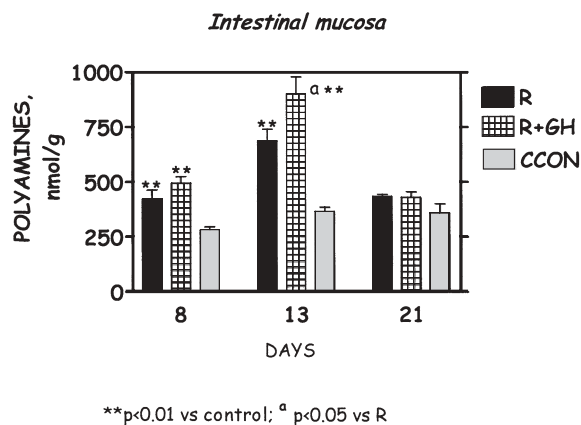


**Fig. 2.** Red blood cell polyamine concentrations during intestinal adaptation; the effect of growth hormone therapy. R, resection; R + GH, resection and growth hormone treatment; CCon, bowel transection control; Control, intact rats

exception of those belonging to R (resected) 21 days group, indicating that the adaptive growth of intestinal remnant is mirrored by circulating polyamine levels. No significant differences were noted between hrGH-treated and untreated operated rats, although the former tended to have higher polyamine concen-



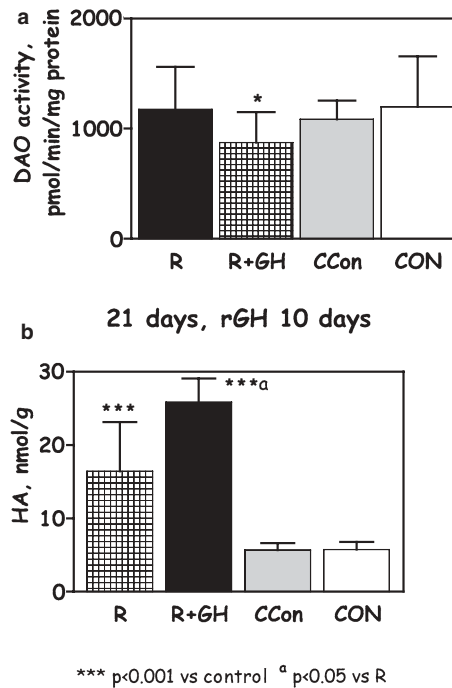
**Fig. 3.** Changes in the activity of ornithine decarboxylase, the enzyme synthesizing putrescine, polyamine precursor, in intestinal mucosa after massive small bowel resection; the effect of growth hormone treatment. *R*, resection; *R + GH*, resection and growth hormone treatment; *CCON*, bowel transection control; *con*, intact rats



**Fig. 4.** Mucosal polyamine concentration in rat small intestine adapting after massive bowel resection; the effect of GH therapy. *R*, resection; *R + GH*, resection and growth hormone treatment; *CCON*, bowel transection control

trations. *R + hrGH* 21 values remained significantly different from the control.

Changes in the activity of ornithine decarboxylase, the enzyme synthesizing putrescine, polyamine precursor, in the intestinal mucosa after massive small bowel resection are depicted in Fig. 3. The additive stimulating effect of hrGH on ODC increases in the intestinal remnant was only seen during the early regeneration phase. Thereafter the two operated groups did not differ. The increases in the tissue ODC activities were followed by an increase in polyamine concentrations as illustrated in Fig. 4. hrGH treated rats have significantly ( $p < 0.05$ ) higher polyamine concentrations than the untreated counterparts on 13<sup>th</sup> day, following the ODC peak. However, weeks after the



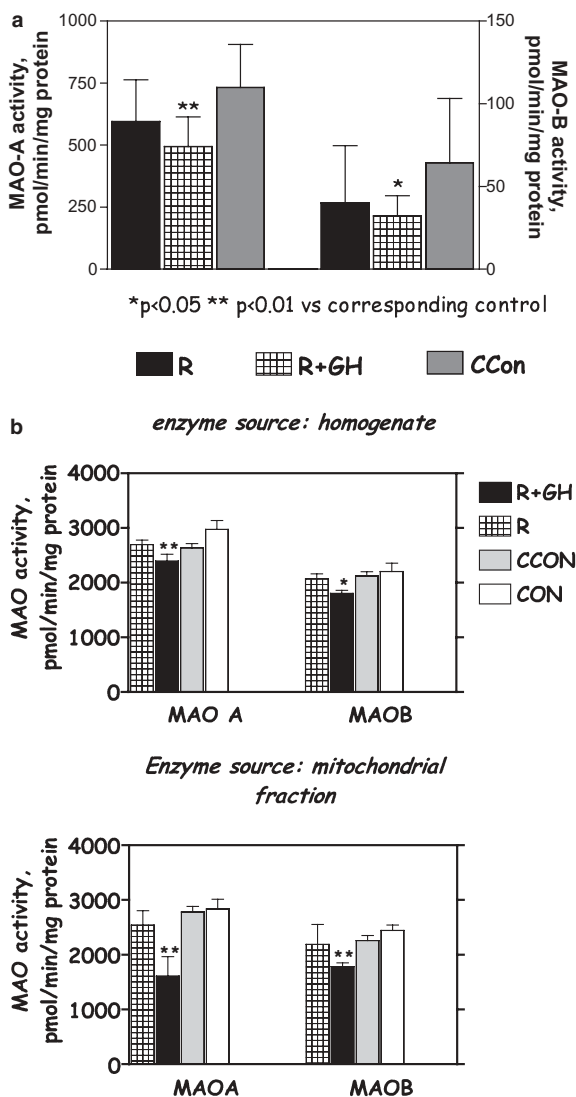
**Fig. 5.** Adaptational processes in rat small bowel after massive resection; the effect of growth hormone therapy. Panel **a**: diamine oxidase activity. Panel **b**: tissue histamine concentration. *R*, resection; *R + GH*, resection and growth hormone treatment; *CCON*, bowel transection control; *CON*, intact rats

surgery the polyamine levels were similar in all groups. Spermidine was the main polyamine in the intestinal mucosa, accounting for  $>90\%$  ( $93.1 \pm 0.07\%$ ) of total tissue polyamines in normal rat small bowel and slightly but not significantly less ( $88.3 \pm 0.13$ ) in regenerating bowel as measured on day 13<sup>th</sup> post surgery.

Growth hormone treatment had the opposite effect on the putrescine catabolizing enzyme, diamine oxidase, activity. Thus, operated rats that were treated with growth hormone for 10 days had significantly lower DAO activities on the 21<sup>st</sup> day post surgery, when regeneration processes were already accomplished (Fig. 5a). Reduced DAO activity in the intestinal mucosa coincided with a higher tissue concentration of histamine, the other DAO substrate (Fig. 5b). These animals also had significantly reduced MAO A and B activities in both the intestinal mucosa (Fig. 6a) and liver (Fig. 6b).

## Discussion

Several studies have shown that growth hormone stimulates growth of different organs, including the



**Fig. 6.** Activities of monoamine oxidase A (MAO A) and B (MAO B) in after massive small bowel resection; the effect of growth hormone therapy. Panel **a**: intestinal mucosa; Panel **b**: liver. R, resection; R + GH, resection and growth hormone treatment; CCon, bowel transection control; CON, intact rats

small intestine (Kaplan, 1990; Leblond and Carriere, 1955; Lehy et al., 1986). Commercial availability of recombinant human growth hormone has triggered interest in its use as a therapeutic measure to support adaptive growth of the intestine after massive small bowel resection. The results of clinical and experimental studies that employed animal model of short bowel syndrome on the effect of GH therapy are inconsistent. Both positive effects and no effects in enhancing the morphological intestinal adaptation have been described (Shulman et al., 1992; Byrne et al., 1995; Liu et al., 1996; Socha et al., 1996; Scolapio

et al., 1997; Vanderhoof et al., 1997; Ljungmann et al., 2000). However, it is well established that induction of ornithine decarboxylase, the key enzyme in polyamine synthesis, is a common cellular response to any trophic stimulus and therefore may be used as an index of the trophic response (Janne et al., 1978; Hosomi et al., 1987). In the present study, by monitoring ornithine decarboxylase activity (Fig. 3) at different periods following the resection and GH therapy we observed that at early stage of adaptation, growth hormone added positively to ODC peak with no further stimulatory effect on the enzyme activity later on. This higher ODC activity in hormone treated resected rats as compared to GH untreated counterparts could be regarded as acceleration of adaptative growth of the bowel remnant. It corresponded with the higher mucosal DNA concentration, although the latter did not attain statistical significance when compared with resected GH untreated rats.

We have shown here that intestinal hyperplasia triggered by massive gut resection was reflected by a significant increase in circulating polyamines, justifying use of erythrocyte polyamine concentration measurement as the marker of small bowel proliferative activity. However, growth hormone treatment did not further enhance it to a great extent, treated rats only tended to have higher RBC polyamine levels even if the intestinal polyamine concentration was significantly higher on day 13 (Fig. 2 and Fig. 4).

The finding that hormone treated animals expressed significantly lower activities of the amine oxidative deamination enzymes was unexpected. Taking into account that diamine oxidase activity reflects the maturational state of enterocytes (Baylin et al., 1978; Luk et al., 1980), the lower diamine oxidase activities in GH treated rats (Fig. 5a) may suggest that there are more less differentiated enterocytes in the intestinal epithelium of these rats. Although specific receptors are present in GI tract (Lobie et al., 1990), most of the enterotrophic effects of GH are thought to be mediated by insulin-like growth factor I (IGF-I) (Behringer et al., 1990; Ohneda et al., 1997). A comparison of IGF-1 transgenic mice (Ohneda et al., 1997) with those over expressing GH (Ulshen et al., 1993), led Ohneda et al. (1997) to conclude that while the effects on intestinal length and mass of both factors were similar, IGF-1 stimulates differentiation of intestinal epithelial cells less effectively. Thus, IGF-1 mediation might be a plausible explanation of reduced DAO activity. Of the FAD dependent amine

oxidases, monoamine oxidase A and B, MAO A is evenly expressed in mature and dividing cells but MAO B appears to be lower in crypt cells (Fogel and Maslinski, 1992). So while similar reasoning could be applied to MAO B, it does not explain lower MAO A activity in resected rats receiving hrGH. Moreover, the effect on MAO enzymes was not restricted to small bowel; it was also observed in the liver. Unlike cytosolic diamine oxidase, monoamine oxidases are expressed in mitochondrial outer membranes (Ramsay, 1998). Growth hormone effects on membrane lipid distribution (Clejan and Maddaiah, 1986) might affect the catalytic activities of monoamine oxidase enzymes (Buckman et al., 1983).

While in some cases the alteration in oxidative deamination of amines may produce unwanted reactions e.g. cheese reaction, in some others as for example in depression, could be of benefit.

It is tempting to speculate that GH exerts its effects on appetite, cognitive functions, memory, mood and other brain functions (Nyberg, 2000) at least in part by interfering with biogenic amine metabolism at the level of amine oxidases. However, no measurements were done in this study on brain tissue.

## Acknowledgement

We thank Ms Krystyna Adach, Zofia Grzelinska, Ing. M.Sc, and Ms Halina Kuchciak for skillful technical assistance.

## References

- Baylin SB, Stevens SA, Shakir KMM (1978) Association of diamine oxidase and ornithine decarboxylase with maturing cells in rapidly proliferating epithelium. *Biochem Biophys Acta* 541: 415–419
- Behringer RR, Lewin TM, Quaife CJ, Palmiter RD, Brinster RL, D'Ercole AJ (1990) Expression of insulin-like growth factor I stimulates normal somatic growth in growth hormone-deficient transgenic mice. *Endocrinology* 127: 1033–1040
- Buckman TD, Eiduson S, Sutphin MS, Chang R (1983) Selective effects on catalysis by the multiple forms of monoamine oxidase produced by interactions of acidic phospholipids with mitochondrial membranes. *J Biol Chem* 258: 8670–8676
- Burton K (1968) Determination of DNA content with diphenylamine. *Methods in enzymology*. In: Grossman L, Moldave K (eds) Part B, vol. 12. Academic Press, New York, pp 163–166
- Byrne TA, Persinger RL, Young LS, Ziegler RT, Wilmore DW (1995) A new treatment for patients with short-bowel syndrome: growth hormone, glutamine and a modified diet. *Ann Surg* 222: 243–254
- Clejan S, Maddaiah VT (1986) Growth hormone and liver mitochondria: effects on phospholipid composition and fatty acyl distribution. *Lipids* 21: 677–683
- Ellegard L, Bosaeus I, Nordgren S, Bengtsson BA (1997) Low-dose recombinant human growth hormone increases body weight and lean body mass in patients with short bowel syndrome. *Ann Surg* 225: 88–96
- Fogel WA (1988) Enzymatic histamine catabolism in vertebrate ontogenesis. A comparative study. *Comp Biochem Physiol* 89C: 355–360
- Fogel WA, Ulatowska M, Adach K, Osinska Z (1985) A sum of 14C putrescine metabolites as a measure of DAO activity. *Column chromatography assay*. *Agents Actions* 16: 99–101
- Fogel WA, Maslinski C (1992) Distribution and plausible functions of amine oxidases in the mucosa of mammalian gastrointestinal tract. *Life Sci* 10: 3–11
- Fowler CJ, Tipton KF (1982) Deamination of 5-hydroxytryptamine by both forms of monoamine oxidase by the rat brain. *J Neurochem* 38: 733–736
- Hosomi M, Stace NH, Lirussi F, Smith SM, Murphy GM, Dowling RH (1987) Role of polyamines in intestinal adaptation in the rat. *Eur J Clin Invest* 17: 375–385
- Janne J, Poso H, Raina A (1978) Polyamines in rapid growth and cancer. *Biochim Biophys Acta* 473: 241–293
- Kaplan SA (1990) Growth and growth hormone: disorders of the anterior pituitary. In: Kaplan SA (ed) *Clinical pediatric endocrinology*. WB Saunders, Philadelphia, pp 1–62
- Kobayahi Y (1963) Determination of histidine decarboxylase by liquid scintillation counting of  $^{14}\text{CO}_2$ . *Anal Biochem* 5: 284–290
- Leblond CP, Carriere R (1955) The effect of growth hormone and tyroxine on the mitotic rate of the intestinal mucosa of the rat. *Endocrinology* 56: 261–266
- Lehy T, Accary JP, Dubrasquet M, Lewin JM (1986) Growth hormone-releasing factor (somatocrinin) stimulates epithelial cell proliferation in the rat digestive tract. *Gastroenterology* 90: 646–653
- Liu YW, Liu W, Jiang M (1996) Effect of recombinant human growth hormone on the intestinal structure of rats receiving bowel resection and parenteral nutrition. *Clinical Nutrition* 15: 303–305
- Ljungmann K, Grofte T, Kissmeyer-Nielsen P, Flyvbjerg A, Vilstrup H, Tygstrup N, Laurberg S (2000) GH decreases hepatic amino acid degradation after small bowel resection in rats without enhancing bowel adaptation. *Am J Physiol Gastrointest Liver Physiol* 279: G700–G706
- Lobie PE, Beipohl S, Waters MJ (1990) Growth hormone receptor expression in the rat gastrointestinal tract. *Endocrinology* 126: 299–306
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951) Protein measurement with the Folin phenol reagent. *J Biol Chem* 193: 265–275
- Luk GD, Bayless TM, Baylin SB (1980) Diamine oxidase (histaminase) a circulating marker for rat intestinal mucosal maturation and integrity. *J Clin Invest* 66: 66–70
- Mates JM, Marquez M, Garcia-Caballero M, Nunez de Castro I, Sanchez-Jimenez (1992) Simultaneous fluorometric determination of intracellular polyamines separated by reversed-phase high-performance liquid chromatography. *Agents Actions* 36: 17–21
- Matsumoto T, Furuta T, Nimura Y, Suzuki O (1982) Increased sensitivity of the fluorometric method of Snyder and Hendley for oxidase assays. *Biochem Pharmacol* 31: 2207–2209
- Nyberg F (2000) Growth hormone in the brain: characteristics of specific brain targets for the hormone and their functional significance. *Frontiers in Neuroendocrinology* 21: 330–348
- Moulinoux J-Ph, Quemener V, Chevet D (1981) Red cell free polyamine concentrations in patients on maintenance hemodialysis. *Life Sci* 29: 955–962

- Moulinoux J-Ph, Quemener V, Quash GA (1984) In vitro studies on the entry of polyamines into normal red blood cells. *Biochimie* 66: 385–393
- Moulinoux J-Ph, Quemener V, Khan NA (1991) Biological significance of circulating polyamines in oncology. *Cell Mol Biol* 37: 773–783
- Ohneda K, Ulshen MH, Fuller CR, D'Ercole AJ, Lund PK (1997) Enhanced growth of small bowel in transgenic mice expressing human insulin-like growth factor I. *Gastroenterology* 112: 444–454
- Peterson CA, Ney DM, Hinton PS, Carey V (1996) Beneficial effect of insuline-like growth factor I on epithelial structure and function in parenterally fed rat jejunum. *Gastroenterology* 111: 1501–1508
- Ramsay RR (1998) Substrate regulation of monoamine oxidases. *J Neural Transm [Suppl 52]*: 139–147
- Scolapio JS, Camilleri M, Fleming CR, Oenning LV, Burton DD, Sebo TJ, Batts KP, Kelly DG (1997) Effect of growth hormone, glutamine and diet on adaptation in short bowel syndrome: a randomized, controlled study. *Gastroenterology* 113: 1074–1081
- Seidel ER (1986) Hormonal regulation of postprandial induction of gastrointestinal ornithine decarboxylase activity. *Am J Physiol* 251(4 Pt 1): G460–466
- Shulman DI, Hu CS, Duckett G, Lavalley-Grey M (1992) Effects of short-term growth hormone therapy in rats undergoing 75% small intestinal resection. *J Pediatr Gastroenterol Nutr* 14: 3–11
- Socha J, Książyk J, Fogel WA, Kierkus M, Lyszkowska, Sasiak K (1996) Is growth hormone a feasible adjuvant in the treatment of children after small bowel resection? *Clinical Nutrition* 15: 185–188
- Ulshen MH, Dowling RH, Fuller CR, Zimmermann EM, Lund PK (1993) Enhanced growth of small bowel in transgenic mice overexpressing bovine growth hormone. *Gastroenterology* 104: 973–980
- Vanderhoof JA, Langnas AN, Pinch LW, Thompson JS, Kaufman SS (1992) Short bowel syndrome. *J Pediatr Gastroenterol Nutr* 14: 359–370
- Vanderhoof JA, Kollman KA, Griffin S, Adrian TE (1997) Growth hormone and glutamine do not stimulate intestinal adaptation following massive small bowel resection in the rat. *J Pediatr Gastroenterol Nutr* 25: 327–331

---

**Authors' address:** Prof. W. Agnieszka Fogel, Institute of Biogenic Amines, Polish Academy of Sciences, 90364 Lodz, Poland, Fax: +48 42 6815283, E-mail: wafogel@mazurek.man.lodz.pl